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Causal role of immune cells in schizophrenia: Mendelian randomization (MR) study



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Abstract

Background Complex immune-brain interactions that affect neural development, survival and function might have causal and therapeutic implications for psychiatric illnesses. However, previous studies examining the association between immune inflammation and schizophrenia (SCZ) have yielded inconsistent findings.

Methods Comprehensive two-sample Mendelian randomization (MR) analysis was performed to determine the causal association between immune cell signatures and SCZ in this study. Based on publicly available genetic data, we explored causal associations between 731 immune cell signatures and SCZ risk. A total of four types of immune signatures (median fluorescence intensities (MFI), relative cell (RC), absolute cell (AC), and morphological parameters (MP)) were included. Comprehensive sensitivity analyses were used to verify the robustness, heterogeneity, and horizontal pleiotropy of the results.

Results After FDR correction, SCZ had no statistically significant effect on immunophenotypes. It was worth mentioning some phenotypes with unadjusted low *P*-values, including FSC-A on NKT (β =0.119, 95% *CI*=0.044 ~ 0.194, *P*=0.002), DN (CD4-CD8-) NKT %T cell (β =0.131, 95% *CI*=0.054 ~ 0.208, *P*=9.03 × 10⁻⁴), and SSC-A on lymphocytes (β =0.136, 95% *CI*=0.059 ~ 0.213, *P*=5.43 × 10⁻⁴). The causal effect of SCZ IgD on transitional was estimated to 0.127 (95% *CI*=0.051 ~ 0.203, *P*=1.09 × 10⁻³). SCZ also had a causal effect on IgD+ %B cell (β =0.130, 95% *CI*=0.054 ~ 0.207, *P*=8.69 × 10⁻⁴), and DP (CD4⁺CD8⁺) %T cell (β =0.131, 95% *CI*=0.054 ~ 0.207, *P*=8.05 × 10⁻⁴). Furthermore, four immunophenotypes were identified to be significantly associated with SCZ risk: naive CD4⁺ %T cell (*OR*=0.986, 95% *CI*=0.979 ~ 0.992, *P*=1.37 × 10⁻⁵), HLA DR on CD14⁻ CD16⁻ (*OR*=0.738 (95% *CI*=0.642 ~ 0.849, *P*=2.00 × 10⁻⁵), CD33^{dim} HLA DR⁺ CD11b⁻ AC (*OR*=0.631, 95% *CI*=0.529 ~ 0.753, *P*=3.40 × 10⁻⁷) and activated & resting Treg % CD4 Treg (*OR*=0.937, 95% *CI*=0.906 ~ 0.970, *P*=1.96 × 10⁻⁴).

Conclusions Our study has demonstrated the close connection between immune cells and SCZ by genetic means, thus providing guidance for future clinical research.

Keywords Schizophrenia, Immunity, Causal inference, Brain, MR analysis, Sensitivity

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Introduction

Schizophrenia (SCZ) is a serious psychiatric disorder that often presents with hallucinations, delusions, and extremely disorganized thinking and behavior, which can affect daily physical functioning and may eventually lead to disability [1, 2]. Global Burden of Disease (GBD) research has demonstrated that there are approximately 20.9 million patients with schizophrenia worldwide [3]. At present, antipsychotic medications are the main method of SCZ treatment [4]. SCZ patients often need lifelong treatment, so early diagnosis and treatment may help to control symptoms before serious complications and improve the prognosis [5, 6].

Epidemiologic studies have demonstrated that earlylife infection is linked with autoimmune disease and subsequent mental disorders in adults [7-10]. Complex immune-brain interactions that affect neural development, survival, and function might have causal and therapeutic implications for disorders including psychiatric illness [1, 11-13]. Cytokines play an important role in infection and inflammation and are crucial mediators of the crosstalk between the brain and the immune system. SCZ leads to a decrease in T helper type 1 (Th1) and an increase in T helper type 2(Th2) cytokine secretion, which are associated with an imbalance in inflammatory cytokines [14]. Proinflammatory cytokines, which are produced at the site of infection by activated accessory immune cells, leading to endocrine, autonomic and behavior changes, include interleukin-1 α and β (IL- 1α and IL-1 β), tumor necrosis factor- α (TNF- α) and interleukin-6 (IL-6) [12]. A systematic review revealed increased soluble interleukin 2 receptor (SIL-2R), and IL-6 and decreased interleukin-2 (IL-2) in SCZ patients, with no significant differences in other cytokines, compared with the general population [14]. However, findings concerning the association between immune inflammation and SCZ have been inconsistent thus far, possibly due to limited sample sizes, flaws in study design, and confounding factors beyond the scope of the existing studies [15-18].

Mendelian randomization (MR) is an analytical method mainly used in epidemiological etiology inference that is based on Mendelian independent distribution law. It is of great importance that the causal sequence of MR is reasonable [19, 20]. Previous observational studies have found many associations between immune cell traits and SCZ, proving the hypothesis of a correlation between them [21, 22]. In this study, a comprehensive two-sample MR analysis was performed to determine the causal association between immune cell signatures and SCZ.

Materials and methods Study design

We assessed the causal relationship between 731 immune cell signatures (7 groups) and schizophrenia based on a two-sample MR analysis. MR uses genetic variation to represent risk factors, and therefore, valid instrumental variables (IVs) in causal inference must satisfy three key assumptions: (1) genetic variation is directly associated with exposure; (2) genetic variation is not associated with possible confounders between exposure and outcome; and (3) genetic variation does not affect outcome through pathways other than exposure. The studies included in our analysis were approved by the relevant institutional review boards, and participants provided informed consent.

Genome-wide association study (GWAS) data sources for SCZ

GWAS summary statistics for SCZ were obtained from the Psychiatric Genomics Consortium (PGC) [23]. The study performed a GWAS on 150,064 European individuals ($N_{case} = 36,989$, $N_{control} = 113,075$), with approximately 9.5 million variants analyzed after quality control and imputation. This GWAS identified 128 independent single nucleotide polymorphisms (SNPs) (83 not previously reported), including at least 108 independent genomic loci under the level of genome-wide significance ($P < 5 \times 10^{-8}$).

Immunity-wide GWAS data sources

GWAS summary statistics for each immune trait are publicly available from the GWAS Catalog (accession numbers from GCST0001391 to GCST0002121) [22]. A total of 731 immunophenotypes including absolute cell (AC) counts (n=118), median fluorescence intensities (MFI) reflecting surface antigen levels (n=389), morphological parameters [MP] (n=32) and relative cell (RC) counts (n=192) were included. Specifically, the MFI, AC and RC features contain B cells, CDCs, mature stages of T cells, monocytes, myeloid cells, TBNK (T cells, B cells, natural killer cells), and Treg panels, while the MP feature contains CDC and TBNK panels. The original GWAS on immune traits was performed using data from 3,757 European individuals and there was no overlapping cohorts. Approximately 22 million SNPs genotyped with high-density arrays were imputed with Sardinian sequence-based reference panel [24] and associations were tested after adjusting for covariates (i.e., sex, age and age^2).

Selection of instrumental variables (IVs)

In accordance with recent research [22, 25], the significance level of IVs for each immune trait was set to 1×10^{-5} . The clumping procedure in PLINK software

(version v1.90) was used to prune these SNPs (linkage disequilibrium [LD] r^2 threshold < 0.1 within 500 kb distance) [26], where LD r^2 was calculated based on 1000 Genomes Projects as a reference panel. For SCZ, we adjusted the significance level to 5×10^{-8} . The proportion of phenotypic variation explained (PVE) and F statistic were calculated for each IV to evaluate IV strength and avoid weak instrumental bias. A total of 7 to 1,786 independent IVs for immunophenotype were determined and these generated IVs could explain an average of 0.240% (range 0.004 – 3.652%) of the variance in their respective immune traits. Then, 108 IVs for SCZ were preserved for further analysis, after removing IVs with low F statistics (<10).

Statistical analysis

All analyses were performed in R 3.5.3 software (http://www.Rproject.org).

To evaluate the causal association between 731 immunophenotypes and SCZ, inverse variance weighting (IVW) [27], weighted median-based [28] and mode-based methods [29] were mainly performed by using the 'Mendelian-Randomization' package (version 0.4.3) [30]. Cochran's Q statistic and corresponding *p* values were used to test the heterogeneity among selected IVs. If the null hypothesis is rejected, random effects IVW was used instead of fixed-effects IVW [27]. To exclude the effect of horizontal pleiotropy, a common method was used (i.e., MR-Egger), which implies the presence of horizontal multiplicity if its intercept term is significant [31]. Furthermore, a powerful method, the MR pleiotropy residual sum and outlier (MR-PRESSO) method was utilized to exclude possible horizontal pleiotropic outliers that could substantially affect the estimation results in the MR-PRESSO package [32]. In addition, scatter plots and funnel plots were used. Scatter plots showed that the results were not affected by outliers. Funnel plots demonstrated the robustness of the correlation and no heterogeneity.

Results

Exploration of the causal effect of SCZ onset on immunophenotypes

To explore the causal effects of SCZ on immunophenotypes, two-sample MR analysis was performed, and the IVW method was used as the main analysis. After multiple test adjustment based on the FDR method, no immune trait was identified at a significance of 0.05. At the significance of 0.20, 7 suggestive immunophenotypes were identified, of which 2 were in the B cell panel, 1 in the cDC panel and 4 in the TBNK panel. We found that SCZ onset could increase the level of *FSC-A* on *NKT* (β =0.119, 95% CI=0.044~0.194, *P*=0.002, *P*_{*FDR*} = 0.185, Fig. 1, Supplementary Tables 1, 2). *DN* (*CD4-CD8-*) *NKT* %*T cells* were increased in SCZ patients

 $(\beta = 0.131, 95\% \text{ CI} = 0.054 \sim 0.208, P = 9.03 \times 10^{-4}, P_{FDR} =$ 0.155, Fig. 1, Supplementary Tables 1, 2). SSC-A on lym*phocytes* was also found to be increased (β =0.136, 95%) CI=0.059~0.213, $P=5.43\times10^{-4}$, $P_{FDR} = 0.155$, Fig. 1, Supplementary Tables 1, 2). The causal effect of SCZ on IgD on transitional was estimated to be 0.127 (95% CI=0.051~0.203, $P=1.09\times10^{-3}$, $P_{FDR} = 0.155$, Fig. 1, Supplementary Tables 1, 2), but the weighted median did not support this association: weighted median (β =0.096, 95% CI = $-0.015 \sim 0.206$, P=0.089). For IgD+ %B cell, a positive association was observed (β =0.130, 95% CI=0.054~0.207, $P=8.69\times10^{-4}$, $P_{FDR} = 0.155$, Supplementary Table 2), which was consistent the with weighted median and MR-PRESSO, but inconsistent with weighted the mode. Similar associations were found for DP $(CD4^+CD8^+)$ %T cell (β =0.131, 95% CI=0.054~0.207, $P = 8.05 \times 10^{-4}$, $P_{FDR} = 0.155$, Supplementary Table 2), and CD11c⁺monocyte AC (β =0.124, 95% CI=0.046~0.202, P=0.002, $P_{FDR} = 0.188$, Supplementary Table 2). The results of the other three methods and sensitivity analysis proved the robustness of the causal associations observed (Supplementary Tables 1, 3). Specifically, the intercept of MR-Egger and global test of MR-PRESSO ruled out the possibility of horizontal pleiotropy (Supplementary Table 1). Scatter plots and funnel plots also indicated the stability of the results (Supplementary Fig. 1 and Fig. 3).

Exploration of the causal effect of immunophenotypes on SCZ

After FDR adjustment (P_{FDR} <0.05), we detected protective effects of four immunophenotypes on schizophrenia: naive CD4+%T cell (maturation stages of T cell panel), HLA DR on CD14⁻CD16⁻ (monocyte panel), CD33^{dim}HLA DR⁺CD11b⁻AC (myeloid cell panel) and activated & resting Treg % CD4 Treg (Treg panel). Specifically, the odds ratio (OR) of naive CD4⁺%T cell on SCZ risk was estimated to be 0.986 (95% CI=0.979~0.992, $P=3.97\times10^{-6}$, $P_{FDR} = 0.004$, Supplementary Table 4) by using the IVW method. Similar results were observed by using three more methods: weighted mode (OR=0.984, 95% CI=0.973~0.995, P=0.005); weighted median (OR=0.986, 95% CI=0.977~0.995, P = 0.003); and MR-PRESSO (OR=0.986, 95% CI=0.979~0.992, $P=2.14\times10^{-5}$). The OR of HLA DR on CD14⁻CD16⁻ on SCZ risk was estimated to be 0.738 (95% CI=0.642~0.849, $P=2.00\times10^{-5}$, $P_{FDR} = 0.005$, Supplementary Table 4) by using the IVW method. Similar results were observed by using three more methods: weighted mode (OR=0.744, 95% CI=0.621~0.891, P = 0.001);weighted median (OR=0.714, 95% CI=0.581~0.878, P = 0.001);and MR-PRESSO CI=0.631~0.860, (OR=0.737, 95% $P=2.50\times10^{-4}$). The OR of CD33^{dim}HLA DR⁺CD11b⁻AC on SCZ risk was estimated to be 0.631 (95% CI=0.529~0.753,

Traits	Methods			OR (95%CI)	Р	P _{FDR}
FSC-A on NKT	IVW (fixed)		Hel	0.119 (0.044, 0.194)	0.002	0.185
	IVW (random)		Hel	0.119 (0.044, 0.194)	0.002	
	Weighted mode	•		0.252(-0.027, 0.531)	0.076	
	Weighted median		HEH	0.149(0.041, 0.257)	0.007	
	MR-Egger (slope)			0.07(-0.24, 0.379)	0.660	
	MR-Eager (intercept)			0.004 (-0.02, 0.028)	0.746	
	MR-PRESSO (raw)			0.073(-0.005, 0.020)	0.070	
DN (CD4-CD8-) NKT %T cell	IVW (fixed)		Hel	0.131 (0.054, 0.208)	9.03×10 ⁻⁴	0.155
	IVW (random)		Hel	0.131 (0.054, 0.208)	9.03×10 ⁻⁴	
	Weighted mode			0.269(-0.029, 0.566)	0.077	
	Weighted median		HIH	0.184 (0.073, 0.294)	0.001	
	MR-Egger (slope)			0.056(-0.262, 0.374)	0.728	
	MR-Egger (intercent)			0.056(-0.262, 0.374)	0.720	
	MR-PRESSO (raw)	1		0.008 (-0.019, 0.031)	0.030	
SSC-A on lymphosyte			Land Land	0.1 (0.024, 0.175)	5.42×10-4	0 155
SSC-A on lymphocyte			1	0.136 (0.059, 0.213)	5.43×10	0.155
	Vivighted mede			0.136 (0.059, 0.213)	5.43×10*	
	vveighted mode			0.305 (0.008, 0.601)	0.044	
	Vveighted median		HEH	0.192 (0.082, 0.303)	6.28×10-4	
	MR-Egger (slope)			0.023 (-0.295, 0.341)	0.889	
	MR-Egger (intercept)			0.009 (-0.016, 0.034)	0.470	
	MR-PRESSO (raw)		H=-1	0.097 (0.021, 0.173)	0.014	
IgD on transitional	IVW (fixed)		H	0.127 (0.051, 0.203)	1.09×10 ⁻³	0.155
	IVW (random)		HEH	0.127 (0.051, 0.203)	1.09×10 ⁻³	
	Weighted mode			-0.018 (-0.273, 0.238)	0.892	
	Weighted median			0.096 (-0.015, 0.206)	0.089	
	MR-Egger (slope)			-0.006 (-0.316, 0.304)	0.969	
	MR-Egger (intercept)			0.011 (-0.013, 0.035)	0.386	
	MR-PRESSO (raw)		HEH	0.089 (0.01, 0.168)	0.030	
lgD+ %B cell	IVW (fixed)		Hel	0.13 (0.054, 0.207)	8.69×10 ⁻⁴	0.155
	IVW (random)		Hel	0.13 (0.054, 0.207)	8.69×10 ⁻⁴	
	Weighted mode	H		0.152 (-0.121, 0.425)	0.276	
	Weighted median		Hel	0.158 (0.047, 0.269)	0.005	
	MR-Egger (slope)	⊢ −∎		-0.049 (-0.364, 0.266)	0.759	
	MR-Egger (intercept)			0.014 (-0.01, 0.039)	0.249	
	MR-PRESSO (raw)		HEH	0.098 (0.017, 0.179)	0.020	
DP (CD4+CD8+) %T cell	IVW (fixed)		Hel	0.131 (0.054, 0.207)	8.05×10 ⁻⁴	0.155
	IVW (random)		Hel	0.131 (0.054, 0.207)	8.05×10 ⁻⁴	
	Weighted mode	H		0.183 (-0.102, 0.468)	0.208	
	Weighted median		HeH	0.136 (0.025, 0.247)	0.016	
	MR-Egger (slope)			-0.051 (-0.364, 0.263)	0.751	
	MR-Egger (intercept)			0.015 (-0.01, 0.039)	0.242	
	MR-PRESSO (raw)		Herl	0.098 (0.02, 0.177)	0.016	0.188
CD11c+ monocyte AC	IVW (fixed)		Hel	0.124 (0.046, 0.202)	0.002	
	IVW (random)		Hel	0.124 (0.046, 0.202)	0.002	
	Weighted mode	—		-0.049 (-0.261, 0.164)	0.653	
	Weighted median			0.093 (-0.015, 0.201)	0.091	
	MR-Egger (slope)			-0.091 (-0.414, 0.233)	0.583	
	MR-Egger (intercept)			0.017 (-0.008. 0.043)	0.181	
	MR-PRESSO (raw)			0.118 (0.053. 0.183)	5.57×10 ⁻⁴	
		-1 0	כ	1		

Fig. 1 Forest plots showed the causal associations between SCZ and immune cell traits. IVW: inverse variance weighting; CI: confidence interval

 $P=3.40\times10^{-7}$, $P_{FDR} = 2.32\times10^{-4}$, Supplementary Table 4) by using the IVW method. Similar results were observed by using three more methods: weighted mode (OR=0.630, 95% CI=0.467~0.849, P=0.002); weighted median (OR=0.562, 95% CI=0.431~0.732, $P=1.95\times10^{-5}$; and MR-PRESSO (OR=0.631, 95%) CI=0.527~0.757, $P=1.28\times10^{-6}$). The OR of activated & resting Treg % CD4 Treg on SCZ risk was estimated to be 0.937 (95% CI=0.906~0.970, $P=1.96\times10^{-4}$, $P_{FDR}=$ 0.034, Supplementary Table 4) by using the IVW method. Similar results were observed by using three more methods: weighted mode (OR=0.945, 95% CI=0.899~0.994, P = 0.028);weighted median (OR = 0.928,95% CI=0.879~0.978, P=0.006); MR-PRESSO (OR=0.937, 95% CI=0.908~0.967, $P=1.48\times10^{-3}$). Additionally, both the intercept of MR-Egger and the global test of MR-PRESSO ruled out the possibility of horizontal pleiotropy

for these four associations. Detailed information from the sensitivity analysis proved the robustness of the causal associations observed (Supplementary Tables 5, Fig. 2). Scatter plots and funnel plots also indicated the stability of the results (Supplementary Fig. 2).

Discussion

Based on large publicly available genetic data, we explored causal associations between 731 immune cell traits and SCZ. To our knowledge, this is the first MR analysis to explore the causal relationship between multiple immunophenotypes and SCZ. In this study, among four types of immune traits (MFI, RC, AC and MP), SCZ was found to have causal effects on seven immunophenotypes (FDR<0.20), and four immunophenotypes had significant causal effects on SCZ (FDR<0.05).

Methods	OR (95% CI)		Р	P _{FDR}
Naive CD4+ %T cell*				
IVW (fixed)	0.986 (0.98, 0.992)		3.97×10 ⁻⁶	0.004
IVW (random)	0.986 (0.979, 0.992)		1.37×10⁻⁵	
Weighted mode	0.984 (0.973, 0.995)	H	0.005	
Weighted median	0.986 (0.977, 0.995)		0.003	
MR-Egger (slope)	0.985 (0.978, 0.993)		1.64×10 ⁻⁴	
MR-PRESSO	0.986 (0.979, 0.992)	•	2.14×10 ⁻⁵	
HLA DR on CD14- CD16-*				
IVW (fixed)	0.738 (0.642, 0.849)		2.00×10 ⁻⁵	0.005
IVW (random)	0.738 (0.636, 0.858)		7.20×10⁻⁵	
Weighted mode	0.744 (0.621, 0.891)	—	1.28×10 ⁻³	
Weighted median	0.714 (0.581, 0.878)	⊢	1.37×10⁻³	
MR-Egger (slope)	0.752 (0.639, 0.886)	⊢	1.16×10⁻³	
MR-PRESSO	0.737 (0.631, 0.86)		2.50×10 ⁻⁴	
CD33dim HLA DR+ CD11b- AC*				
IVW (fixed)	0.631 (0.529, 0.753)		3.40×10 ⁻⁷	2.32×10 ⁻⁴
IVW (random)	0.631 (0.526, 0.757)		7.40×10 ⁻⁷	
Weighted mode	0.63 (0.467, 0.849)	← ∎───┤	0.002	
Weighted median	0.562 (0.431, 0.732)	←■───┤	1.95×10⁻⁵	
MR-Egger (slope)	0.622 (0.495, 0.782)	<-∎	6.30×10 ⁻⁵	
MR-PRESSO	0.631 (0.527, 0.757)	⊢ −■−−1	1.28×10 ⁻⁶	
Activated & resting Treg % CD4 Tre	eg			
IVW (fixed)	0.937 (0.906, 0.97)	HEH	1.96×10 ⁻⁴	0.034
IVW (random)	0.937 (0.906, 0.97)	HEH	1.96×10 ⁻⁴	
Weighted mode	0.945 (0.899, 0.994)	H=-1	0.028	
Weighted median	0.928 (0.879, 0.978)	⊢ ∎-1	0.006	
MR-Egger (slope)	0.959 (0.907, 1.014)	⊢ ∎-	0.141	
MR-PRESSO	0.937 (0.908, 0.967)	HeH	1.48×10 ⁻³	
		0.5 1	1.2	

Fig. 2 Forest plots showed the causal associations between immune cell traits and SCZ by using different methods. IVW: inverse variance weighting; CI: confidence interval

Our study found that the risk of SCZ decreased with an increase in the proportion of naive CD4+%T cell. Naive CD4+% T cells are able to modulate proinflammatory and anti-inflammatory signals by differentiating into a variety of T-helper (Th) cell lineages, each with its own distinct cytokine profile and function. There is evidence that secreted cytokines play a role in the occurrence and progression of SCZ [33, 34]. Compared with healthy people, IL-6 and TNF- α levels were significantly increased in schizophrenia patients, while IL-2, IL-4 and IFN- γ levels were significantly decreased [35].

HLA DR on CD14-CD16- in the monocyte panel has been proven to be associated with decreased SCZ risk [36, 37]. HLA-DR is an MHC class II cell surface receptor encoded by the human leukocyte antigen complex on chromosome 6 region 6P21. Previous studies have shown that lower concentrations of IL-1 β in cells may reflect the weakening of monocyte function in SCZ [38]. The low expression of HLA-DR in monocytes in chronic inflammation demonstrates the anti-inflammatory effect of HLA-DR molecules.

Activated and resting Treg% CD4 Tregs have also been proven to be significantly associated with a reduced risk of SCZ [39, 40]. Regulatory T cells (Tregs) are the key immunomodulatory cells involved in the control of inflammatory processes, and their function is directly related to the human leukocyte antigen (HLA) gene, which has been implicated in genetic studies of SCZ. A significantly increased proportion of Tregs in patients with SCZ compared to healthy controls. Tregs were found to be able to improve the negative symptoms of schizophrenia by offsetting the ongoing inflammatory process [41].

In addition, it was noteworthy that the presence of SCZ was found to be associated with increased FSC-A on NKT, DN (CD4-CD8-) NKT %T cell, SSC-A on lymphocytes, IgD on transitional cells, IgD+ %B cell, DP (CD4+CD8+) %T cell, and CD11c+monocyte AC levels. Several studies have found increased concentrations of inflammatory cytokines in the blood of patients with SCZ [42]. To date, there has been limited research on the role of NKT cells in neurological diseases, and a casecontrol study found an increase in the relative number of NK cells in schizophrenia [43]. Notably, Finkelstein et al. reported that the function of NKT cells in neurodegeneration is limited [44]. Mechanistically, the regulation of NKT cells induces a cytokine shift in the liver and promotes the recruitment of T cells into the affected spinal cord [45].

This study conducted two-sample MR analysis based on the published results of large GWAS cohorts, with a large sample size of approximately 150,000 people, so it has high statistical efficiency. The conclusions of this study are based on genetic instrumental variables, and causal inference is made using a variety of MR analysis methods. The results are robust and were not confounded by horizontal pleiotropy and other factors. Our study has limitations. First, even when multiple sensitivity analyses are performed, horizontal pleiotropy cannot be fully assessed. Second, due to the lack of individual information, we cannot conduct further stratified analysis of the population. Third, the study was based on a European database, so the conclusion cannot be extended to other ethnic groups, which would limit the generalizability of our results. Finally, we used a looser threshold to evaluate the results, which may increase some falsepositives while simultaneously enabling a more comprehensive assessment of the strong association between the immune profile and SCZ.

Conclusions

In conclusion, we have demonstrated the causal associations between several immunophenotypes and SCZ through a comprehensive bidirectional MR analysis, highlighting the complex pattern of interactions between the immune system and SCZ.

Furthermore, our research significantly reduced the impact of unavoidable confounding factors, reverse causality, and other factors. It may provide a new avenue for researchers to explore the biological mechanisms of SCZ and can lead to exploration of earlier intervention and treatment. Our results extend the findings of psychoimmunology, providing valuable clues for the prevention of SCZ.

List of Abbreviations

AC	Absolute cell
CI	Confidence interval
FDR	False discovery rate
GBD	Global Burden of Disease
GWAS	Genome-wide association study
HLA	Human leukocyte antigen
IL-1α	Interleukin-1a
IL-1β	Interleukin-1β
IL-2	Interleukin-2
IL-6	Interleukin-6
IVs	Instrumental variables
IVW	Inverse variance weighting
LD	Linkage disequilibrium
MFI	Median fluorescence intensities
MP	Morphological parameters
MR	Mendelian Randomization
MR-PRESSO	MR pleiotropy residual sum and outlier
OR	Odds ratio
PGC	Psychiatric Genomics Consortium
PVE	Phenotypic variation explained
RC	Relative cell
SCZ	Schizophrenia
SNPs	Single Nucleotide Polymorphisms
TNF-α	Tumor necrosis factor-α

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12888-023-05081-4.

Supplementary file 1: Causal effects of schizophrenia on immune cells

Supplementary file 2: IVW results of the causal effect of SCZ on immune cells

Supplementary file 3: Results of the causal effect of immune cells on SCZ

Supplementary file 4: Sensitivity analysis results of causal effects of SCZ on immune cells

Supplementary file 5: Sensitivity analysis results of causal effects of immune cells on SCZ

Supplementary file 6: Supplementary figures

Supplementary file 7: Supplementary Methods

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Authors' contributions

ZYS and YZ contributed to conceptualization, supervision and writing review and editing. CDW contributed to formal analysis and writing—original draft. DDZ, DJZ, XWZ, LY, TL and XDG contributed to acquisition of data. CLH contributed to statistical analysis. All authors contributed to the article and approved the submitted version.

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Data Availability

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation. Further inquiries can be directed to the corresponding author.

Declarations

Ethics approval and consent to participate

All participants signed informed consent to study protocols approved by the Sardinian Regional Ethics Committee (protocol no. 2171/CE). All studies were approved by local ethics committees, and all participants provided written, informed consent. Informed consent was obtained from all participants and/ or their LAR. This study was conducted in accordance to relevant guidelines and regulations.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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